

Supplementary Information

for

Flow cytometry-based quantification of targeted knock-in events in human cell lines using a GPI-anchor biosynthesis gene *PIGP*

Md. Lutfur Rahman, Toshinori Hyodo, Muhammad Nazmul Hasan, Yuko Mihara,
Sivasundaram Karnan, Akinobu Ota, Shinobu Tsuzuki, Yoshitaka Hosokawa, and
Hiroyuki Konishi*

*Department of Biochemistry, Aichi Medical University School of Medicine,
Nagakute, Aichi 480-1195, Japan*

* Corresponding author

Hiroyuki Konishi

Department of Biochemistry, Aichi Medical University School of Medicine

1-1 Yazako Karimata, Building #2, Room 362,

Nagakute, Aichi 480-1195, Japan

Phone: +81-561-62-3311 ext. 12362

Fax: +81-561-61-4056

E-mail: hkonishi@aichi-med-u.ac.jp

- **Supplementary Figures 1 and 2**
- **Supplementary Tables 1 and 2**

Supplementary Figure 1

Donor-*PiGPex3*

-174 #1 -154 #2 -127 -75
 --aacgagcta gaagaactga aaggagaaa aaatcatttc tgagtaatc agggacactt caaaagagct tttctgttt atctaatct taatagattt
 --tttgctcgat cttcttgact ttccctccccc tttagtaaag actcaatttgc tccctgtgaa gttttctcgaa aaagacaaaa tagacttaga attatctaacc

-74
 atagatttata tgtagtgcttt actgacttctt ctccctcttt tttttttttt tcttttctac agTACTTTAC CTCGTGTGGG CCTTTATTCC TGAATCTTGG
 tatctaatct acatacagaa tgactgaaga gaaggagaaa aaaaagaaaa agaaaagatg tcATGAAATG GAGCACACCC GGAAATAAGG ACTTAGAACC
 I L Y L **V** **W** A F I P E S W

+26
 CTAAACTCTT TAGGTTAAC CTATTGGCCT CAAAAttagt ttaatatat ttcttaatg agatataat taatgctgtt ttgaggagtc atatcatttc--
 GATTTGAGAA ATCCAAATTG GATAACCGGA GTTTcatc aaattatata aagaatttac tccttatata attacgacaa aactccttag tatagtaaag--
 L N S L G L T Y W P Q K

#4 +82 +125

Genomic region surrounding *PiGP* exon 3

-174 #1 -154 #2 -127 -75
 --aacgagcta gaagaactga aaggagaaa aaatcatttc tgagtaatc agggacactt caaaagagct tttctgttt atctaatct taatagattt
 --tttgctcgat cttcttgact ttccctccccc tttagtaaag actcaatttgc tccctgtgaa gttttctcgaa aaagacaaaa tagacttaga attatctaacc

-74
 atagatttata tgtagtgcttt actgacttctt ctccctcttt tttttttttt tcttttctac agTACTTTAC CTC**TAGA**GGG CCTTTATTCC TGAATCT**TGG**
 tatctaatct acatacagaa tgactgaaga gaaggagaaa aaaaagaaaa agaaaagatg tcATGAAATG GAG**ATC**CCC GGAAATAAGG ACTTAGAACC
 I L Y L * **R** A F I P E S W

+26
 CTAAACTCTT TAGGTTAAC CTATTGGCCT CAAAAttagt ttaatatat ttcttaatg agatataat taatgctgtt ttgaggagtc atatcatttc--
 GATTTGAGAA ATCCAAATTG GATAACCGGA GTTTcatc aaattatata aagaatttac tccttatata attacgacaa aactccttag tatagtaaag--
 L N S L G L T Y W P Q K

#4 +82 +125

Supplementary Figure 1 DNA sequences surrounding the knock-in site at *PiGP* exon 3 in the Donor-*PiGPex3* plasmid (top) and the DLD-1-derived *PiGP* reporter clone (bottom). Target sequences of a Cas9 nuclease and Cas9 nickases are shaded in blue and green, respectively, with darker shading on their neighboring protospacer adjacent motifs (PAMs). Numbers with plus or minus signs near Cas9 target sites indicate the distance from the knock-in site (± 0) to theoretical cleavage sites within individual Cas9 target sites. Nucleotides shown by red and green bold letters represent a *PiGP* truncating mutation in the DLD-1-derived *PiGP* reporter clone and its corresponding wild-type sequence in Donor-*PiGPex3*, respectively. Upper case and lower case letters indicate exonic and intronic sequences, respectively.

Supplementary Figure 2

Donor-*PIGPex2*

-134 #5 -93 **-35**
--gtacccgcgg cgagcgagg aggcgagaat ggatcaatgg tgccacggag cacatcgctg acgctgattt ttttcctttt ccacagATTG TCTAAAGCCC
--catggcgcc gcctcgctcc tccgctcta cctagttacc acgggtgcctc gtgttagcgtac tgcgactaac acaaggaaaa ggtgtcAAC AGATTCGGG
-34 #6 **+65**
CAGGAAAAAT GGTGGAAAAT TCACCGTCGC CAT**GG**CAGA AAGAGCGATT TATGGCTTTG TTCTTTCTT AAGCTCCAA TTTGGCTTCAGA gtaagtactt
GTCCTTTTA CCACCTTTA AGTGGCAGCG GT**ACGG**TCT TTCTCGCTAA ATACCGAAC AAGAAAAGAA TTCGAGGGTT AAACCGAAGT cattcatgaa
M V E N S P S P **L** P E R A I Y G F V L F L S S Q F G F
+66 #8 +118 **+165**
tcctgatgga tggccagggg cttttctgg cgttcagtga caagggaga atacat**tgg**t tagactgtgc acacatgaac ttttgattct ttctccat--
aggactacct acc**gg**tcccc gaaaagacc gcaagtcaacttccat tatgtaaaccatctgacacg tttgtacttg acaactaaga aagaggata--
+85 #7

Genomic region surrounding *PIGP* exon 2

-134 #5 -93 **-35**
--gtacccgcgg cgagcgagg aggcgagaat ggatcaatgg tgccacggag cacatcgctg acgctgattt ttttcctttt ccacagATTG TCTAAAGCCC
--catggcgcc gcctcgctcc tccgctcta cctagttacc acgggtgcctc gtgttagcgtac tgcgactaac acaaggaaaa ggtgtcAAC AGATTCGGG
-34 F -6 ±0 **+65**
CAGGAAAAAT GGTGGAAAAT TCACCGTCGC CAT**GG**TAAGA AAGAGCGATT TATGGCTTTG TTCTTTCTT AAGCTCCAA TTTGGCTTCAGA gtaagtactt
GTCCTTTTA CCACCTTTA AGTGGCAGCG GT**ACCA**TCT TTCTCGCTAA ATACCGAAC AAGAAAAGAA TTCGAGGGTT AAACCGAAGT cattcatgaa
M V E N S P S P **W** * E R A I Y G F V L F L S S Q F G F
+66 #8 +118 **+165**
tcctgatgga tggccagggg cttttctgg cgttcagtga caagggaga atacat**tgg**t tagactgtgc acacatgaac ttttgattct ttctccat--
aggactacct acc**gg**tcccc gaaaagacc gcaagtcaacttccat tatgtaaaccatctgacacg tttgtacttg acaactaaga aagaggata--
+85 #7

Supplementary Figure 2 DNA sequences surrounding the knock-in site at *PIGP* exon 2 in the Donor-*PIGPex2* plasmid (top) and the HCT116-derived *PIGP* reporter clone (bottom). For annotations of the numbers and colors shown in this figure, see the legend for Supplementary Figure 1.

Supplementary Table 1: Cas9 target sites with PAMs

| Target name | Sequence |
|--|-----------------------------------|
| <i>Creation of the initial deletion</i> | |
| <i>PIGP</i> -nuclease-A | ACTGTTGTGAGGATTAAC <u>GGGG</u> |
| <i>PIGP</i> -nuclease-B | AAGGGATGACTAGTTCACAT <u>TGG</u> |
| <i>Extension of the deletion in HCT116</i> | |
| <i>TTC3</i> -nuclease-C | TAAATCCATAGGCTGACAC <u>AGGG</u> |
| Junction-nuclease-D | CTGTTGTGAGGATTAACATT <u>GGG</u> |
| <i>PIGP</i> correction assay using the DLD-1-derived <i>PIGP</i> reporter clone | |
| <i>PIGP</i> -nickase-1 | GAGCTAGAAGAACTGAA <u>AGGGAGG</u> |
| <i>PIGP</i> -nickase-2 | AATCATTCTGAGTTAAC <u>CTAGGG</u> |
| <i>PIGP</i> -nickase-3 | GGGCCTTATTCTGAAT <u>CTTGG</u> |
| <i>PIGP</i> -nickase-4 | TTTAATATATTCTAA <u>ATGAGG</u> |
| <i>PIGP</i> -nuclease-E | TACAGTAC <u>TTTACCTCTAGAGGG</u> |
| <i>PIGP</i> correction assay using the HCT116-derived <i>PIGP</i> reporter clone | |
| <i>PIGP</i> -nickase-5 | AGAATGGATCAATGGTGCC <u>ACGG</u> |
| <i>PIGP</i> -nickase-6 | AGCGTCAGCGATGTGCTCC <u>GTGG</u> |
| <i>PIGP</i> -nickase-7 | TGAACGCCAGAAAAAGCCC <u>CTGG</u> |
| <i>PIGP</i> -nickase-8 | GTGACAAGGGAAGAAC <u>ATACATTGG</u> |
| <i>PIGP</i> -nuclease-F | TGGAAAATT <u>CACCGTCGCCATGG</u> |

PAM, protospacer adjacent motif

Underlining indicates PAM sequences.

Supplementary Table 2: PCR primers generated in this study

| Primer name | Sequence |
|--|---|
| <i>Creation of dual sgRNA vectors</i> | |
| U6 promoter-F | 5'-CCTAGGT <u>CTAGAGAGGGCCTATTCCCATGATTCC</u> |
| CAG-R | 5'-CCATTACCGTAAGTTATGTAAC |
| <i>Creation of Donor-PIGPex3</i> | |
| PIGPint2-F2 | 5'-CTTTAG <u>AGACTCATCGTGAGCACTTGTAA</u> |
| PIGPint3-R | 5'-TTT <u>CTGGATCCTGCATTATCTCTATTCCCTT</u> |
| <i>Creation of Donor-PIGPex2</i> | |
| TTC3ex1-R | 5'-CTCGG <u>CTGCAGCGGGACAA</u> |
| PIGPint2-R1 | 5'-CCTACCC <u>CATAAGAACTGTTCATGCGTC</u> |
| <i>Analytical PCRs probing deletions</i> | |
| PIGPint2-F1 | 5'-TGGCTGACTTC <u>ACTCTGCTG</u> |
| PIGPint2-R2 | 5'-AGCTT <u>CTAACATCCTGACATACTCTT</u> |
| PIGPdownstream-F1 | 5'-CAGGGT <u>GAGGCAACTAAGCA</u> |
| PIGPdownstream-R1 | 5'-CCAGC <u>CTACAAAGGCCCTCA</u> |
| TTC3int1-F | 5'-GCCAC <u>CTGGCTCTCTTGT</u> |
| TTC3int1-R | 5'-CAC <u>CTCACCTGCCACTAAAG</u> |
| <i>Production of PIGP Southern probe</i> | |
| PIGPdownstream-F2 | 5'-CTGAG <u>TTTGGTCAAGCAGGG</u> |
| PIGPdownstream-R2 | 5'-ACACTAA <u>ACTGATCATGACCCTCA</u> |

PCR, polymerase chain reaction

Underlining indicates the restriction enzyme sites used for the incorporation of PCR products into a plasmid.