

## Supplementary legends

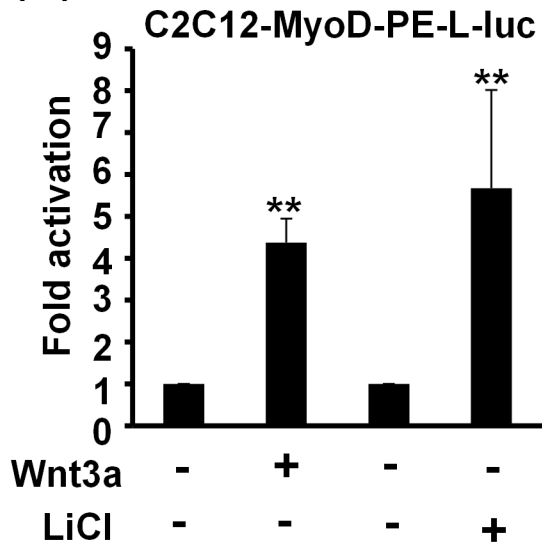
### **FIGURE S1**: LiCl activates MyoD-PE-L-luc dose-dependently.

C2C12-MyoD-PE-L-luc stable clones were treated with Wnt3a or LiCl (10 mM) and their luciferase activity was determined 48 hr after treatment (A). Their response to increasing amount (2, 5, and 10 mM) of LiCl was also determined (B). \*\*:  $p < 0.01$  vs. vehicle control.

### **FIGURE S2** Pax7 protein levels in C2C12 cells.

The levels of Pax7 protein in total lysate (50  $\mu$ g) from C2C12 cells of CMB and MT stages were examined with Western blot using antibodies against Pax7 and Gapdh. Numbers to the left represent molecular weight in kilodalton (Kd).

(A)



(B)

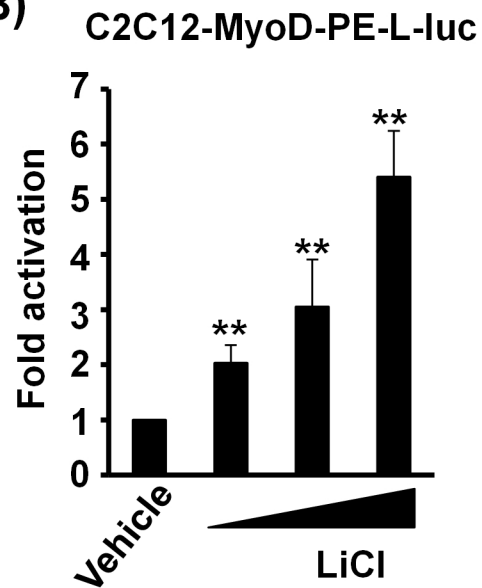
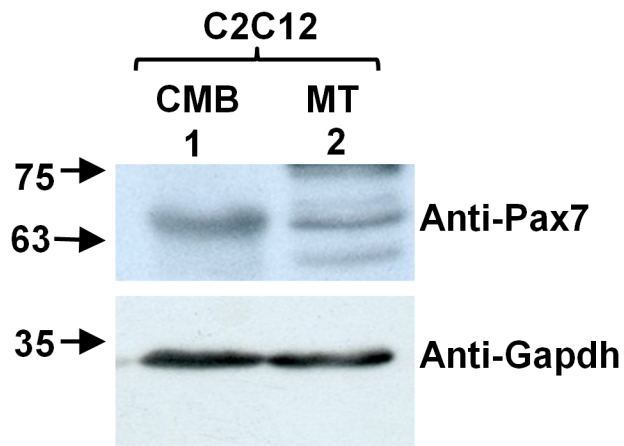


FIGURE S2



**Table S1: Primer sets used for cloning.**

NCU code	Clone name	Primer Sequence*	Amplicon size (bp)
026022 026023	<i>DE</i>	FP : 5'- <b>cggaattc</b> gtgttgcccagttatcccag-3' RP : 5'- <b>cggaattc</b> gctggctgggtctctctagtagtac-3'	4497
021003 021004	<i>P6P</i>	FP : 5'- <b>ggggtacc</b> ccagtagaggcaaagaaaggaa-3' RP : 5'- <b>ctagctag</b> ctaggtgggttggaatgcacac-3'	2965
026022 039005	<i>DE-#1</i>	FP : 5'- <b>cggaattc</b> gtgttgcccagttatcccag-3' RP : 5'- <b>cggaattc</b> cgctgcctgcctcaggcct-3'	1776
039004 039010	<i>DE-#2</i>	FP : 5'- <b>cggaattc</b> gctcttcacctcttc-3' RP : 5'- <b>cggaattc</b> gcatgctcactctggc-3'	1489
039002 039003	<i>DE-#3</i>	FP : 5'- <b>cggaattc</b> gccaggcccttccttct-3' RP : 5'- <b>cggaattc</b> gagtgacagagacggcc-3'	1491
039011 026023	<i>DE-#4</i>	FP : 5'- <b>cggaattc</b> gagcccaggggtacatgaa-3' RP : 5'- <b>cggaattc</b> gctggctgggtctctagtagtac-3'	1215
026022 039010	<i>DE-#5</i>	FP : 5'- <b>cggaattc</b> gtgttgcccagttatcccag-3' RP : 5'- <b>cggaattc</b> gcatgctcactctggc-3'	2581
026022 039003	<i>DE-#6</i>	FP : 5'- <b>cggaattc</b> gtgttgcccagttatcccag-3' RP : 5'- <b>cggaattc</b> gagtgacagagacggcc-3'	3269
039002 026023	<i>DE-#7</i>	FP : 5'- <b>cggaattc</b> gccaggcccttccttct-3' RP : 5'- <b>cggaattc</b> gctggctgggtctctagtagtac-3'	2712
039004 039003	<i>DE-#8</i>	FP : 5'- <b>cggaattc</b> gctcttcacctcttc-3' RP : 5'- <b>cggaattc</b> gagtgacagagacggcc-3'	2177
039004 026023	<i>DE-#9</i>	FP : 5'- <b>cggaattc</b> gctcttcacctcttc-3' RP : 5'- <b>cggaattc</b> gctggctgggtctctagtagtac-3'	3398
039004 039023	<i>DE-#10</i>	FP : 5'- <b>cggaattc</b> gctcttcacctcttc-3' RP : 5'- <b>cggaattc</b> gctggtaccaggagg-3'	2922
33005 33006	<i>A fragment</i>	FP : 5'- <b>cgacgcg</b> tcg tat gta agg cta atg aga tcc cgg -3' RP : 5'- <b>cgacgcg</b> tcg ctg gct cag ggt ctg gga -3'	1402
33026 33027	<i>B fragment</i>	FP : 5'- <b>cgacgcg</b> t acttgcagccaagcagccct -3' RP : 5'- <b>cgacgcg</b> t gcagccctgagatccc -3'	1161

33024 33025	<i>C fragment</i>	FP : 5'- <b>cgacgcgt</b> gggctcccagggtataa -3' RP : 5'- <b>cgacgcgt</b> gaaggaaggaaggaggag -3'	1381
33022 33023	<i>D fragment</i>	FP : 5'- <b>cgacgcgt</b> ctctcctgaacactac -3' RP : 5'- <b>cgacgcgt</b> ctccgctcaccccccccc -3'	1316
33020 33021	<i>E fragment</i>	FP : 5'- <b>cgacgcgt</b> caggacacaggcagggca -3' RP : 5'- <b>cgacgcgt</b> acttaggaggtggaggt -3'	1340
26027 26028	<i>F fragment</i>	FP : 5'- <b>cgacgcgtc</b> ccatcatatcatetaagcta -3' RP : 5'- <b>cgacgcgtc</b> ccacaattcccagattctcttgc -3'	1116
33018 33019	<i>G fragment</i>	FP : 5'- <b>cgacgcgt</b> cctgectcctcttttccc -3' RP : 5'- <b>cgacgcgt</b> ccattctgtcccattcc -3'	1366
33016 33030	<i>H fragment</i>	FP : 5'- <b>cgacgcgt</b> gat get ccc aga ctc cca -3' RP : 5'- <b>cgacgcgt</b> aactcaccatgtacc -3'	1254
33014 33015	<i>I fragment</i>	FP : 5'- <b>cgacgcgt</b> ggg tac atg gtg agt gtc -3' RP : 5'- <b>cgacgcgt</b> gtg ggt ggg gtg ggg tgg g -3'	1290
33011 33012	<i>J fragment</i>	FP : 5'- <b>cgacgcgt</b> cccaccccaccccacccac -3' RP : 5'- <b>cgacgcgt</b> accctccaagtatccc -3'	1240
033034 033043	<i>K fragment</i>	FP : 5'- <b>cgacgcgt</b> CCTCCTCTGTGCTGCCTTA-3' RP : 5'- <b>cgacgcgt</b> GGTTTCCTTTCTAAGTGC-3'	715
33007 33008	<i>L fragment</i>	FP : 5'- <b>cgacgcgt</b> gcaggcatgaaggacatg -3' RP : 5'- <b>cgacgcgt</b> ggtgttagtcattctgggg -3'	1510
33003 33004	<i>M fragment</i>	FP : 5'- <b>cgacgcgtcg</b> cct gcc ttt cac acc tga tg -3' RP : 5'- <b>cgacgcgtcg</b> aca cat gtc tca tcc -3'	1174
33001 33002	<i>N fragment</i>	FP: 5'- <b>cgacgcgtc</b> ggt acc tga tgt gtg cct gg -3' RP: 5'- <b>cgacgcgtcg</b> tat gct cta aat aaa atc tat tc -3'	1142
039020 039021	<i>β-Catenin-Δ90</i>	FP: 5'- <b>cgcgatccatggctcagagggtcc</b> -3' RP: 5'- <b>cgcgatcccaggtcagatcaaaccag</b> -3'	2079
039026 039021	<i>β-Catenin-Δ151</i>	FP: 5'- <b>cgcgatccatggcaattcctgagct</b> -3' RP: 5'- <b>cgcgatcccaggtcagatcaaaccag</b> -3'	1896

\*: linkers and restriction sites are shown in bold.

FP: forward primer

RP: reverse primer

**Table S2: Primer sets used for amplifying gene in RT-PCR**

NCU code	Gene name	Primer Sequence	Amplicon size
00324 00325	<i>MyoD</i>	FP : 5'-ggctacgacaccgcctacta-3' RP : 5'-gttctgtgtcgcttaggat-3'	204 bp
006012 006013	<i>Myogenin</i>	FP : 5'-ccagtgaatgcaactcccacagc-3' RP : 5'-agacatatcctccaccgtga-3'	166 bp
001042 001043	<i>Gapdh</i>	FP : 5'-cctctggaaagctgtggcgt-3' RP : 5'-ttggcaggtttctccaggcg-3'	190 bp
006001 006002	<i>M36b4</i>	FP : 5'-ggcagcattataaccctgaagtg-3' RP : 5'-cggacaccctccagaaagc-3'	73 bp

**FP: forward primer**

**RP: reverse primer**

**Table S3: Primer sets used for amplifying genes in CHIP assay**

NCU Code	Name	Primer sequence	Amplicon size (bp)
039047 039048	<i>MyoD-P1</i>	FP: 5'- ccc ctt ccc ctc acc at- 3' RP: 5'- cgg tga gtc cct cca ag - 3'	102
039049 039050	<i>MyoD-P2</i>	FP: 5'- ctt ggt tgt gta tcc tgc t- 3' RP: 5'- tca aat tcc tgt agc c - 3'	160
039051 039052	<i>MyoD -P3</i>	FP: 5'- tga gca agt gtt gcc cag tta t- 3' RP: 5'- gac caa cct cct atg tct a- 3'	121
039041 039042	<i>DKK1</i>	FP: 5'- cca gcg ctt tga aat ccc at - 3' RP: 5'- acc gcg gct gcc ttt ata cc- 3'	100
029124 029125	<i>M-cadherin</i>	FP: 5'-ctc tcc egg gca gcc ctc tg-3' RP: 5'-gcg cat gcg tcc tgc tcc cga t -3'	126
039024 039057	<i>MyoD-L1</i>	FP: 5'- cgg aat tcg cag gca tga agg aca tg - 3' RP: 5'- tgg aca cct cac acg ctc- 3'	187
039058 039059	<i>Id3</i>	FP: 5'- ccg ggc ata cat tta gtt cct - 3' RP: 5'- tct ctc tct cct ctc tct ctc aa- 3'	365

**FP: forward primer****RP: reverse primer**